

# Human Recombinant ADRA1A Adrenoceptors Stable Cell Line

Technical Manual No. TM0429

Version 10132010

I	Introduction .....	1
II	Background.....	1
III	Representative Data.....	2
IV	Thawing and Subculturing.....	3
V	References .....	3
	Limited Use License Agreement.....	4

## I. Introduction

Catalog Number: M00225

Cell Line Name: CHO-K1/ADRA1A

Gene Synonyms:  $\alpha_1$ -adrenergic receptor, ADRA1A

Expressed Gene: Genbank Accession Number NM\_000680; no expressed tags

Host Cell: CHO-K1

Quantity: Two vials of frozen cells ( $3 \times 10^6$  per vial)

Stability: 16 passages

Application: Functional assay for ADRA1A receptor

Freeze Medium: 45% culture medium, 45% FBS, 10% DMSO

Complete Growth Medium: Ham's F12, 10% FBS

Culture Medium: Ham's F12, 10% FBS, 400  $\mu\text{g/ml}$  G418

Mycoplasma Status: Negative

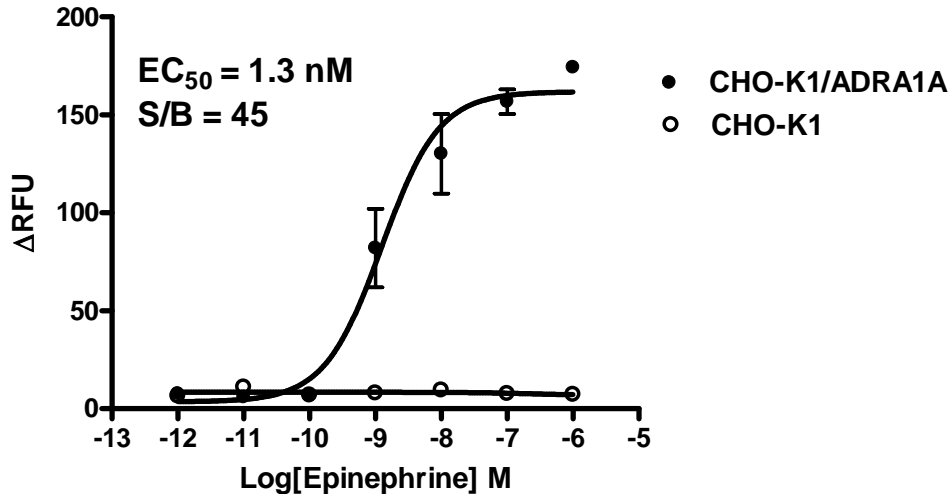
Storage: Liquid nitrogen immediately upon delivery

## II. Background

The  $\alpha_1$ -adrenergic receptor (AR) family consists of three closely related gene products ( $\alpha_{1A}$ ,  $\alpha_{1B}$ , and  $\alpha_{1D}$ ) that mediate the actions of norepinephrine (NE) and epinephrine in sympathetically innervated tissues and brain.  $\alpha_1$ -ARs belong to the G protein-coupled receptor family and consist of single polypeptide chains predicted to have seven transmembrane spanning domains. With similar pharmacological and signaling properties,  $\alpha_1$ -AR subtypes act through Gq/11 proteins to activate phospholipase C, increase both inositol 1,4,5-trisphosphate production and intracellular  $\text{Ca}^{2+}$ . Once activated by binding,  $\alpha_1$ -ARs initiate the cellular pathways leading to the regulation of physiological effects, including blood pressure maintenance, glucose metabolism, renal sodium reabsorption, and cardiac inotropy.

### III. Representative Data

Concentration-dependent stimulation of intracellular calcium mobilization by Epinephrine in CHO-K1/ADRA1A and CHO-K1 cells



**Figure 1.** Epinephrine-induced concentration-dependent stimulation of intracellular calcium mobilization in CHO-K1/ADRA1A and CHO-K1 cells. The cells were loaded with Calcium-4 prior to stimulation with an ADRA1A receptor agonist, Epinephrine. The intracellular calcium change was measured by FlexStation. The relative fluorescent units (RFU) were plotted against the log of the cumulative doses (10-fold dilution) of Epinephrine (Mean  $\pm$  SD, n = 2). The EC<sub>50</sub> of Epinephrine on ADRA1A in CHO-K1 cells was 1.3 nM. The S/B of Epinephrine on ADRA1A in CHO-K1 cells was 45.

Notes:

- EC<sub>50</sub> value is calculated with four parameter logistic equation:  

$$Y = \text{Bottom} + (\text{Top} - \text{Bottom}) / (1 + 10^{((\text{LogEC}_{50} - X) * \text{HillSlope})})$$

X is the logarithm of concentration. Y is the response  
 Y is RFU and starts at Bottom and goes to Top with a sigmoid shape.
- Signal to background Ratio (S/B) = Top/Bottom

### IV. Thawing and Subculturing

Thawing: Protocol

- Remove the vial from liquid nitrogen tank and thaw cells quickly in a 37°C water-bath.
- Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol and transfer the cells to a 15 ml centrifuge tube containing 9 ml of complete growth medium.
- Pellet cells by centrifugation at 200 x g force for 5 min, and discard the medium.
- Resuspend the cells in complete growth medium.
- Add 10 ml of the cell suspension in a 10 cm dish.
- Add G418 to a concentration of 400 µg/ml the following day.

Subculturing: Protocol

1. Remove and discard culture medium.
2. Wash cells with PBS (pH=7.4) to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 ml of 0.05% (w/v) Trypsin- EDTA (GIBCO, Cat No. 25300) solution to 10 cm dish and observe the cells under an inverted microscope until cell layer is dispersed (usually within 3 to 5 minutes).  
Note: To avoid clumping, do not agitate the cells by hitting or shaking the dish while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
4. Add 6.0 to 8.0 ml of complete growth medium and aspirate cells by gently pipetting, centrifuge the cells 200 x g force for 5min, and discard the medium.
5. Resuspend the cells in culture medium and add appropriate aliquots of the cell suspension to new culture vessels.
6. Incubate cultures at 37°C.

Subcultivation Ratio: 1:3 to 1:8 weekly.

Medium Renewal: Every 2 to 3 days

## V. References

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