

**Human Recombinant G<sub>α15</sub> Stable Cell Line**  
**Cat. No. M00554****Version 03232016**

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**I. INTRODUCTION**

Catalog Number: M00554

Cell Line Name: HEK293/G<sub>α15</sub>

Gene Synonyms: GNA15, GNA16

Official Full Name: Guanine nucleotide binding protein (G protein), alpha 15 (Gq class)

Expressed Gene: GenBank Accession Number NM\_002068; no expressed tags

Host Cell: HEK293

Quantity: Two vials of frozen cells (3×10<sup>6</sup> per vial)

Stability: 16 passages

Application: Functional assay for Gs and Gi/o-coupled GPCR receptors

Freeze Medium: 90% FBS, 10% DMSO

Complete Growth Medium: DMEM, 10% FBS

Culture Medium: DMEM, 10% FBS, 0.5 µg/ml puromycin

Mycoplasma Status: Negative

Storage: Liquid nitrogen immediately upon delivery

**II. BACKGROUND**1) HEK293/G<sub>α15</sub>

HEK293/G<sub>α15</sub> is a HEK293 cell line stably expressing the G<sub>α15</sub> alpha subunit protein which a Gq protein. It is used as a host cell for transfection expression of Gs and Gi/o -coupled receptors, the constitutively expressed G<sub>α15</sub> protein in the cells allows many transfected receptors which normally stimulate/inhibit the cAMP pathway, to couple to Gq signal transduction and mobilize intracellular calcium. The cell line carries the puromycin resistance gene and is resistant to puromycin

§: GenScript employs a PCR-based method to test the mycoplasma. The test covers 11 of the most common strains of mycoplasma, (covering approximately 95% of *M. fermentans*, *M. hyorhinis*, *M. arginini*, *M. orale*, *M. salivarium*, *M. hominis*, *M. pulmonis*, *M. arthritis*, *M. neurolyticum*, *M. hyopneumoniae* and *M. capricolum*) and one species *Ureaplasma* (*U. urealyticum*), with sufficient sensitivity and specificity.

**860 Centennial Ave., Piscataway, NJ 08854, USA**Toll-Free: 1-877-436-7274 Tel: 1-732-885-9188 Fax: 1-732-210-0262 Email: [product@genscript.com](mailto:product@genscript.com) Web: [www.genscript.com](http://www.genscript.com)

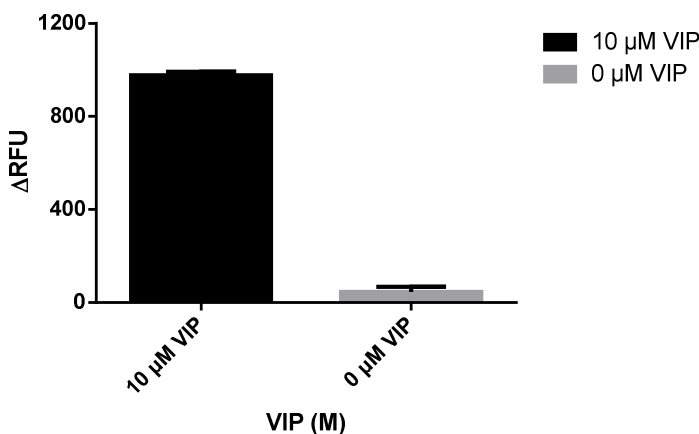
2) The sequence of  $G_{\alpha 15}$

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ATGGCCCCGCTCGCTGACCTGGCGCTGCTGCCCTGGTGCCTGACGGAGGATGAGAAGGCCGCCGCCGGG
TGGACCAGGAGATCAACAGGATCCTCTTGAGCAGAAGAAGCAGGACCGCGGGGAGCTGAAGTGCTGCT
TTTGGGCCCAGGCGAGAGCGGGAAGAGCACCTTCATCAAGCAGATGCGGATCATCCACGGCGCCGGCTAC
TCGGAGGAGGAGCGCAAGGGCTTCCGGCCCCTGGTCTACCAGAACATCTTCGTGTCCATGCGGGCCATGA
TCGAGGCCATGGAGCGGCTGCAGATTCCATTCAGCAGGCCCGAGAGCAAGCACCACGCTAGCCTGGTCAT
GAGCCAGGACCCCTATAAAGTGACCACGTTTGAGAAGCGCTACGCTGCGGCCATGCAGTGGCTGTGGAGG
GATGCCGGCATCCGGGCCTACTATGAGCGTCGGCGGGAATTCCACCTGCTCGATTCAGCCGTGTACTION
TGTCCACCTGGAGCGCATCACCGAGGAGGGCTACGTCACACAGCTCAGGACGTGCTCCGCAGCCGCAT
GCCCACCACTGGCATCAACGAGTACTGCTTCTCCGTGCAGAAAACCAACCTGCGGATCGTGGACGTCGGG
GGCCAGAAGTCAGAGCGTAAGAAATGGATCCATTGTTTCGAGAACGTGATCGCCCTCATCTACCTGGCCT
CACTGAGTGAATACGACCAGTGCCTGGAGGAGAACAACCAGGAGAACCGCATGAAGGAGAGCCTCGCATT
GTTTGGGACTATCCTGGAACCTACCCTGGTTCAAAGCACATCCGTCATCCTCTTTCTCAACAAAACCGAC
ATCCTGGAGGAGAAAATCCCCACCTCCCACCTGGCTACCTATTTCCCCAGTTTCCAGGGCCCTAAGCAGG
ATGCTGAGGCAGCCAAGAGGTTTCATCCTGGACATGTACACGAGGATGTACACCGGGTTCGTGGACGGCCC
CGAGGGCAGCAAGAAGGGCGCACGATCCCGACGCCTCTTCAGCCACTACACATGTGCCACAGACACACAG
AACATCCGCAAGGTCTTCAAGGACGTGCGGGACTCGGTGCTCGCCCCTACCTGGACGAGATCAACCTGC
TGTGA
    
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### III. REPRESENTATIVE DATA

Intracellular calcium mobilization of 10  $\mu$ M VIP on transient-transfected HEK293/ $G_{\alpha 15}$  cells with pcDNA3.1-VPAC2 plasmid



**Figure 1.** Intracellular calcium mobilization of 10  $\mu$ M VIP on transient-transfected HEK293/ $G_{\alpha 15}$  cells with pcDNA3.1-VPAC2 plasmid.

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#### IV. THAWING AND SUBCULTURING

##### Thawing Protocol

1. Remove the vial from liquid nitrogen tank and thaw cells quickly in a 37°C water-bath.
2. Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol and transfer the cells to a 15 ml centrifuge tube containing 9 ml of complete growth medium.
3. Pellet cells by centrifugation at 200 x g force for 5 min, and remove the medium.
4. Re-suspend the cells in complete growth medium.
5. Transfer the cell suspension to a 10 cm dish with 10 ml of complete growth medium.
6. Grow the cells in incubator with 37°C, 5 %CO<sub>2</sub>.
7. In the following day, replace the cells with fresh medium contains antibiotic.

##### Sub-culturing Protocol

1. Remove the culture medium from cells.
2. Wash cells with PBS (pH=7.4) to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 ml of 0.05% (w/v) Trypsin- EDTA (GIBCO, Cat No. 25300) solution into 10 cm dish and observe the cells under an inverted microscope until cell layer is dispersed (usually within 3 to 5 minutes).  
Note: To avoid cells clumping, do not agitate the cells by hitting or shaking the dish while waiting for the cells detach. If cells are difficult to detach, please place the dish in 37°C incubator for ~2 min.
4. Add 6.0 to 8.0 ml of complete growth medium into dish and aspirate cells by gently pipetting.
5. Centrifuge the cells at 200 x g force for 5min, and remove the medium.
6. Resuspend the cells in culture medium and add the cells suspension to new culture dish.
7. Grow the cells in incubator with 37°C, 5 %CO<sub>2</sub>.

Subcultivation Ratio: 1:3 to 1:8 weekly.

Medium Renewal: Every 2 to 3 days

**GenScript USA Inc,**  
860 Centennial Ave.  
Piscataway, NJ 08854  
Toll-Free: 1-877-436-7274  
Tel: 1-732-885-9188, Fax: 1-732-210-0262  
Email: [product@genscript.com](mailto:product@genscript.com)  
Web: <http://www.genscript.com>

**For Research Use Only.**

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