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**Human Recombinant Thyrotropin-releasing Hormone Receptor Stable Cell Line**  
**Cat. No. M00202** **Version 06222020**

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**I. INTRODUCTION**

Catalog Number: M00202

Cell Line Name: CHO-K1/TRH1

Gene Synonyms: MGC141920; TRHR

Expressed Gene: Genbank Accession Number NM\_003301; no expressed tags

Host Cell: CHO-K1

Culture Properties: Adherent

Quantity: Two vials of frozen cells (>1×10<sup>6</sup> per vial)

Stability: More than 16 passages

Application: Functional assay for TRH1 receptor (Calcium flux assay)

Freeze Medium: 45% culture medium, 45% FBS (Cat. #10099-141, Gibco), 10% DMSO (Cat. #D2650, Sigma)

Complete Growth Medium: Ham's F-12K (Kaighn's) (Cat. #21127, Life Technologies), 10% FBS

Culture Medium: Ham's F-12K (Kaighn's), 10% FBS, 400 µg/ml G418 (Cat. #10131-035, Gibco)

Mycoplasma Status: Negative\*

Storage: Liquid nitrogen immediately upon receipt

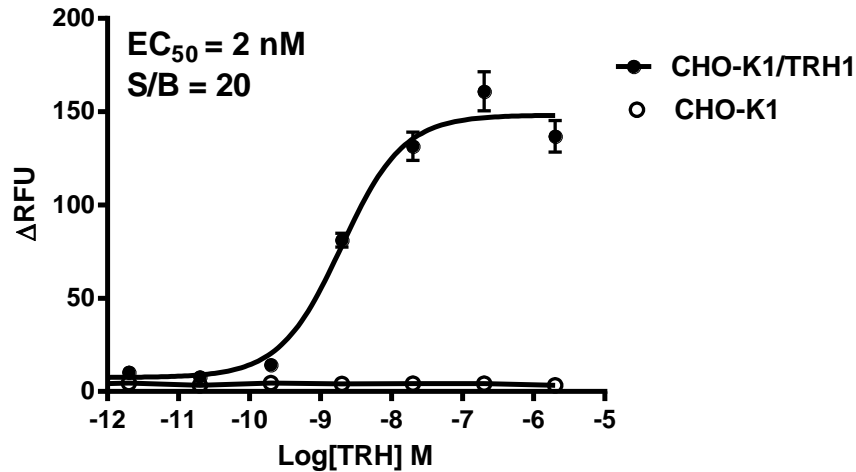
**II. BACKGROUND**

Thyrotropin-releasing hormone receptor1 (TRH1) is a member of G-protein coupled receptor family A. This protein is a receptor for Thyrotropin-releasing hormone (TRH). Human TRH1 is expressed in lymphocytes, pituitary gland and CNS. It can stimulate the releasing of prolactin (PRL), thyrotropin (TSH). TRH1 receptor knockout mice exhibit a slightly reduced growth rate, considerable decrease in serum T<sub>3</sub>, T<sub>4</sub>, and prolactin levels but no alteration of thyroid-stimulating hormone levels.

\* The mycoplasma test was performed with MycoAlert™ PLUS Mycoplasma Detection Kit of Lonza.

### III. REPRESENTATIVE DATA

Concentration-dependent stimulation of intracellular calcium mobilization by TRH in CHO-K1/TRH cells



**Figure 1:** TRH-induced concentration-dependent stimulation of intracellular calcium mobilization in CHO-K1/TRH1 and CHO-K1 cells. The cells were loaded with Calcium-4 prior to being stimulated with a TRH1 receptor agonist, TRH. The intracellular calcium change was measured by FLIPR. The relative fluorescent units (RFU) were normalized and plotted against the log of the cumulative doses (5-fold dilution) of TRH (Mean  $\pm$  SD, n = 2). The EC<sub>50</sub> of TRH on this cells was 2 nM.

**Note:**

- EC<sub>50</sub> value is calculated with four parameter logistic equation:  

$$Y = \text{Bottom} + \frac{(\text{Top} - \text{Bottom})}{(1 + 10^{((\text{LogEC}_{50} - X) * \text{HillSlope}))}}$$
 X is the logarithm of concentration. Y is the response and starts at Bottom and goes to Top with a sigmoid shape.
- Signal to Background Ratio (S/B) = Top/Bottom

### IV. THAWING AND SUBCULTURING

**Thawing Protocol**

- Remove the vial from liquid nitrogen tank and thaw cells quickly in a 37°C water-bath.
- Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol and transfer the cells to a 15 ml centrifuge tube containing 9 ml of complete growth medium.
- Pellet cells by centrifugation at 200 X g for 5 min, and remove the medium.
- Resuspend the cells with 1 ml complete growth medium.
- Transfer the cell suspension to a 10 cm dish containing 10 ml complete growth medium.
- Transfer the dish into an incubator of 37°C, 5% CO<sub>2</sub>.
- Add antibiotic into the medium on the next day.

### Sub-culturing Protocol

1. Remove the culture medium from cells.
2. Wash cells with PBS (pH=7.4) to remove all traces of serum that contains trypsin inhibitor.
3. Add 2.0 ml 0.05% (w/v) Trypsin- EDTA (GIBCO, Cat No. 25300) solution into 10 cm dish and observe the cells under an inverted microscope until cell layer is dispersed (usually within 3 to 5 minutes).

**Note:** To avoid cells clumping, do not agitate the cells by hitting or shaking the dish during incubation. If cells are difficult to detach, please place the dish in 37°C incubator for ~2 min.

4. Add 6.0 to 8.0 ml complete growth medium into dish and aspirate cells by gently pipetting.
5. Centrifuge the cells at 200 X g for 5min, and remove the medium.
6. Resuspend the cells in culture medium and transfer the cells to a new culture dish.
7. Transfer the dish into an incubator of 37°C, 5% CO<sub>2</sub>.

Subcultivation Ratio: 1:3 to 1:8

Medium Renewal: Every 2 to 3 days

### V. REFERENCES

1. Duthie SM (1993) Cloning and functional characterisation of the human TRH receptor. *Mol Cell Endocrinol. Sep*; 95(1-2):R11-5
2. Cao J. Cloning and characterization of a cDNA encoding a novel subtype of rat thyrotropin-releasing hormone receptor. *J. Biol. Chem.*, 273, 32281 - 32287
3. Anderson L. Rapid desensitization of the thyrotropin-releasing hormone receptor expressed in single human embryonal kidney 293 cells. *Biochem J.*, 311, 385 - 392.

**GenScript USA Inc,**  
860 Centennial Ave.  
Piscataway, NJ 08854  
Toll-Free: 1-877-436-7274  
Tel: 1-732-885-9188, Fax: 1-732-210-0262  
Email: [product@genscript.com](mailto:product@genscript.com)  
Web: <http://www.genscript.com>

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