

Human Parathyroid Hormone Receptor 2 Stable Cell Line

Technical Manual No. TM0298

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I. Introduction

The parathyroid hormone receptor 2 (PTH2) is a member of G-protein coupled receptor family 2. This protein is a receptor for parathyroid hormone (PTH). This receptor is more selective in ligand recognition and has a more specific tissue distribution than parathyroid hormone receptor 1 (PTH1). It is activated only by PTH and not by parathyroid hormone-like hormone (PTHLH) and is particularly abundant in the brain and pancreas.

This manual describes establishment of a cell line and a protocol of pharmacologically validated human PTH2 GPCR receptor (Genebank Accession Number: NM_005048).

II. Cell Line Information

- Catalog Number: M00141
- Cell Line Name: HEK293/PTH2/CRE/ β -Lac
- Description:

The HUMAN PTH2 is amplified by PCR using a high fidelity enzyme and subcloned into the pcDNA3.1-hygro (+) mammalian expression vector. The full-length ORF has been confirmed by sequencing. The PTH2 reporter cell line is created by cotransfection of pcDNA3.1-hygro(+)/PTH2 and pCRE/ β -Lac in an HEK293 cell line. The transfected cells are stably selected by 100 μ g/ml hygromycin. Single cell clones with high PTH2 inducibility and low β -lactamase background are isolated using ring cloning. The clones with the largest dynamic ranges in β -lactamase are chosen for pharmacological and stability studies.
- Function: Cell-based, functional assay for PTH2
- Quantity: 2 vial (2×10^6) frozen cells
- Host Cell: HEK293
- Cell Phenotype: Adherent/epithelial
- Recommended Storage: Liquid nitrogen, upon delivery
- Propagation Medium: DMEM, 10% FBS, 100 μ g/ml hygromycin
- Mycoplasma: Negative

III. PTH (1-34) Dose Response of HEK293/PTHR2/CRE/ β -Lac Stable Cell Line and Assay Procedure

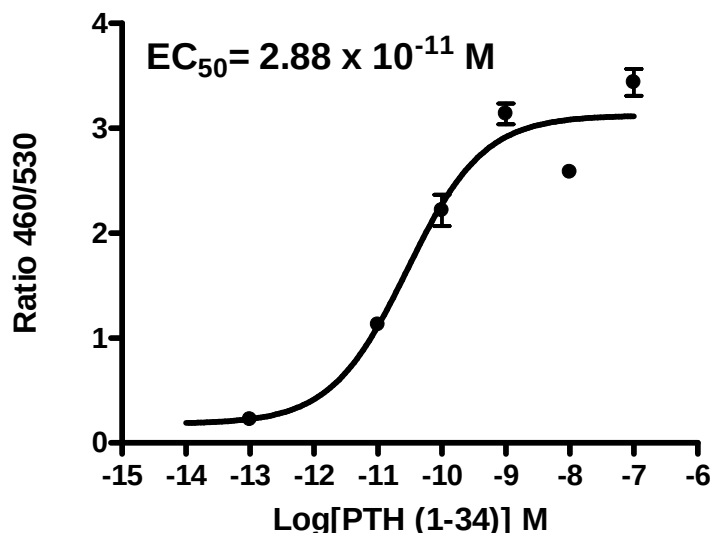


Fig. 1. Dose response of β -lactamase activity as monitored with Analyst HT plate reader upon treatment with ligand. Assay was done according to procedure described as below. Data represent means $\pm$ SEM for triplicate samples. EC_{50} value for PTH (1-34) dose response was determined using GraphPad Prism 4 software.

Assay Procedure

CCF-4 Assay of HEK293/PTHR2/CRE/ β -Lac Reporter Cells

1. Seed 25,000 cells per well in Growth Medium (100 μ L per well) into 96-well tissue culture treated black-wall, clear-bottom plates (Costar #3603) after trypsinization. Prepare some wells with medium alone (no cells) to use for determining plate background.
2. Culture cells in 5% CO_2 at 37°C. Allow cells to reach $\approx$ 90% confluence.
3. 12-24 hours before the assay, replace Growth Medium with 100 μ L/well serum-free DMEM. Be careful not to disturb the cells.
4. Prepare ligand solution in serum-free DMEM (10X).
5. Add 10 μ L of 10X PTH (1-34) solution to wells for stimulation and 10 μ L of serum-free DMEM per well for non-stimulated control.
6. Incubate cells in 5% CO_2 at 37°C for five to six hours.
7. Load cells with 2 μ M CCF4/AM as described in CCF4 Loading Protocol.
8. Incubate the plate at room temperature for 60-120 minutes without shaking.
9. Wait 60-200 minutes after CCF-4 loading. Then either read with Analyst HT plate reader or observe under fluorescence microscope.

CCF-4 Loading Protocol

1. Solution A: Dissolve 5 mg CCF-4/AM with 4.6 ml DMSO to a stock concentration at 1 mM. Aliquot and store at -80°C, protected from light.
2. Solution B: 100 mg/ml Pluronic -F127 surfactant in DMSO and 0.1% acetic acid
3. Solution C: Red Dye solution
4. 6X substrate loading solution:

- 1) Add 6 μ L of Solution A to 60 μ L of Solution B and mix well.
 - 2) Add 934 μ L of Solution C to the combined Solutions A and B and mix well.
 5. Add 6X Substrate Loading Solution to cells to 1X final concentration (e.g., add 20 μ L of 6XCCF4-AM Substrate Loading Solution to 100 μ L of cells in buffer).
 6. Add the same volume of 6X Substrate Loading Solution to the cell-free background control wells containing assay medium or buffer to 1X final concentration.
- Cover the plate to protect it from light and evaporation.

IV. References

1. ALESSANDRO BISELLO *et al.* (2004) Agonist-Specific Regulation of Parathyroid Hormone (PTH) Receptor Type 2 Activity: Structural and Functional Analysis of PTH- and Tuberoinfundibular Peptide (TIP) 39-Stimulated Desensitization and Internalization. 2004. *Molecular Endocrinology* 18: 1486–1498.
2. Thomas J. Gardella and Harald Jüppner. (2001) Molecular properties of the PTH/PTHrP receptor. July 2001, *TRENDS in Endocrinology & Metabolism* Vol.12 No.5

V. Appendix

Cell Culture Conditions

Note: This clone often grows as clumps. It is necessary to separate cells into single cell suspensions whenever the cells are passaged or plated. MATRIGEL Matrix-coated flasks or plates are recommended for healthier cells.

Complete Culture Medium:

DMEM: 90%, FBS: 10%, L-glutamine: 2.0 mM, Amp: 100 μ g/ml, Strep: 100 μ g/ml, hygromycin: 50 μ g/ml.

Freezing Medium:

Growth medium plus 20% FBS and 10% DMSO

Thawing Cells:

1. Quickly thaw frozen cells in a 37°C water bath, agitating continuously.
2. Using a 1 ml pipette, slowly pipet the cells up and down five times and add, drop by drop, to a 15 ml centrifuge tube containing 5 ml of fresh prewarmed complete DMEM medium. Then centrifuge at 1,000 rpm for five minutes.
3. Discard the supernatant medium and resuspend the cell pellet in 5 ml of fresh prewarmed complete DMEM medium. Transfer cells to a T25 flask and incubate at 37°C with 5% CO₂ until the cells reach >90% confluence. The recovery rate for frozen cells is usually 90% or above.

Subculturing:

When the cells reach confluence, they need split. This cell line is normally split twice weekly at 1:8 to 1:15 dilutions.

1. Carefully aspirate all the media. Gently rinse the cell layer with appropriate amount of 0.2% trypsin-EDTA, and aspirate it off.
2. Wait for about 1-3 minutes. Then dislodge the cells by gently tapping the sides of flask or dish.
3. Resuspend cells with appropriate amount of complete DMEM medium, and split cells as desired.

Changing Medium:

This is normally done every other day.

1. Gently aspirate off medium.
2. Transfer fresh warm complete DMEM medium (37°C) into a flask (5 ml for T25 and 10 ml for T75).

Freezing Cells:

1. Repeat the steps 1-3 of subculturing section.
2. Centrifuge down the cells at 1,000 rpm for five minutes.
3. Aspirate off the supernatant and resuspend the cells in fresh freezing medium at a density of $2-3 \times 10^6$ cells/ml. Add 1 ml cells per cryogenic vial.
4. Put the cryogenic vial of cells into cryo freezing container. Then transfer the container to a -80°C environment and leave it there overnight.
5. Transfer cryogenic vial into liquid nitrogen (-196°C).

Reagents & Consumables:

1. DMEM: Dulbecco's Modified Eagle Medium powder, high glucose (Gibco BRL, Cat #12100-046)
2. FBS: Fetal Bovine Serum (Hyclone, Cat #CH30160.03)
3. L-Glutamine: 200 mM (Gibco BRL, Cat # 25030-081)
4. Ampicillin: 50 mg/ml (Sigma A-9518)
5. Streptomycin Sulfate: 50 mg/ml (Gibco BRL, Cat # 11860-038)
6. Hygromycin B in PBS, 50 mg/ml (Invitrogen, Cat #10687-010)
7. Trypsin: 1:250 rom Bovine Pancreas (Gibco BRL, Cat # 27250016)
8. DMSO: dimethyl sulphoxide, for molecular biology (Sigma, Cat #D8418)
9. Hepes: Sigma Cat #H-3375
10. CCF4: (Invitrogen, Cat #K1096)
11. PTH (1-34): synthesized by HD Biosciences
12. Cryogenic Vial: (Corning Cat #430289)
13. 96 Well Plate: Costar, Cat# 3603, Blackwall/clear bottom, Polystyrene, sterilized.

Media and Solutions:

1. PBS (for preparation of 500 ml)
 - 1) KCl: 0.1 g
 - 2) KH_2PO_4 : 0.1 g
 - 3) NaCl: 4.0 g
 - 4) $\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$: 1.4425 g

Dissolve the above components in double-distilled water (ddH_2O) and adjust pH to 7.4 with 0.1 N NaOH. Add ddH_2O to the final volume of 500 ml. Autoclave and store at 4°C.

2. Trypsin-EDTA (for preparation of 100 ml)

- | | |
|-------------|--------|
| 1) Trypsin: | 0.25 g |
| 2) 2%EDTA: | 2 ml |
| 3) PBS: | 98 ml |

Dissolve trypsin in 2%EDTA and PBS completely. Sterilize the solution by passing through a 0.20 µm membrane filter; store at 4°C.

3. Culture medium (for preparation of 1 L)

- 1) Measure out 950 ml distilled water to dissolve the media components, stirring gently until the solution becomes clear.
- 2) Add NaHCO₃ 3.7 g for high glucose DMEM.
- 3) Adjust pH of medium to 0.2-0.3 below the desired final working pH (using 1 N NaOH or 1 N HCL is recommended). Add slowly while stirring.
- 4) Dilute to 1 liter with ddH₂O.
- 5) Sterilize the medium immediately using the method of membrane filtration. Store at 4°C.

4. Ampicillin/streptomycin 50 mg/ml

Dissolve 1 g ampicillin or streptomycin in 20 ml ddH₂O and sterilize the solution by membrane filtration using 0.20 µm filter. Aliquot and store at 4°C for short-term conservation and -20°C for long term conservation.

Map of huHRH2-pCDNA3.1/hygro(+)

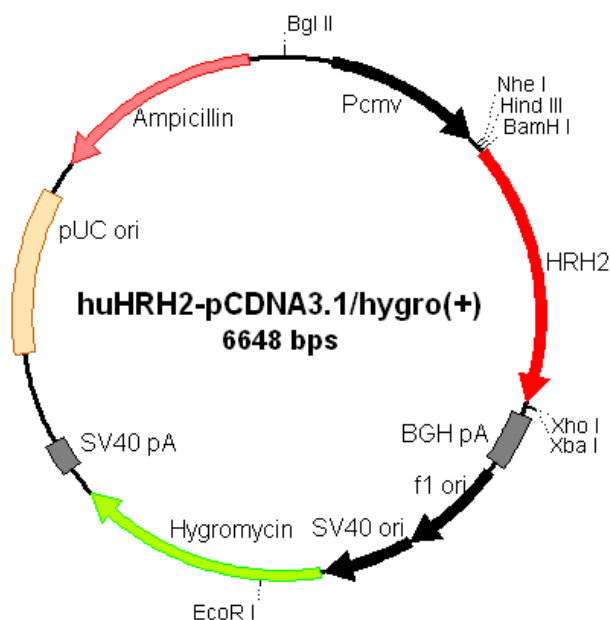
Name: huHRH2-pCDNA3.1/hygro(+)

Insert gene: *Homo sapiens* histamine receptor H2, (HRH2)

Length of insert: 1103bp Sequence reference: Gene Bank NM_022304

Vector: pCDNA3.1/hygro(+) (Invitrogen) Insert site: BamH I (5') and Xho I (3')

Plasmid Map:



Comments for huHRH2-pCDNA3.1/hygro(+) **6648nucleotides**

CMV promoter: bases 232~819
T7 promoter/priming site: bases 863~882
huHRH2 gene: bases 938~2020
pCDNA3.1/BGH reverse priming site: bases 2069~2086
BGH polyadenylation signal: bases 2075~2299
f1 origin: bases 2345~2773
SV40 early promoter and origin: bases 2778~3148
Hygromycin resistance gene (ORF): bases 3166~4191
SV40 early polyadenylation signal: bases 4321~4451
pUC origin: bases 4834~5507 (complementary strand)
Ampicillin resistance gene (bla): bases 5652~6512 (complementary strand)

Name: Human histamine receptor H2 (HRH2) in pCDNA3.1/hygro(+)

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GenScript USA Inc.
120 Centennial Ave., Piscataway, NJ 08854
Tel: 732-885-9188, 732-885-9688
Fax: 732-210-0262, 732-885-5878
Email: info@genscript.com
Web: <http://www.genscript.com>

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Catalog Number: M00141

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